three initial conformational states from X-ray crystal structures is performed. The modes with greatest variance qualitatively agree with the slowest ENM modes, indicating a PCA analysis on the FIRST/FRODA ensemble of conformations produce consistent predictions. However, in order to determine a statistically significant interpretation of the PCA modes, a negative control was performed that replaces a biologically relevant displacement vector with a random displacement vector (decoy). Surprisingly, the decoys produce similar high correlation to the same few greatest variance PCA modes. Our results put serious question on the soundness of the common interpretation of high overlap between a conformational displacement vector and a mode. Further analysis shows that biologically relevant motions found in myosin V require many modes. As such, inferring specific mechanisms based on a single mode is generally difficult to impossible. To facilitate a quantitative justification in identifying a biologically relevant pathway, the protein motion is decomposed further by projecting PCA modes into structural domains, and correlated motions are identified via mode-coupling. We present a critical comparison to recent predictions for the conformational pathways in myosin V, where statistical significance is monitored. This work is supported by NIH grant 1R21HL093531.

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Modeling Drkn Sh3 Domain Using Sequence Specific Wormlike Chain Model

Yujie Chen, William J. Wedemeyer, Lisa J. Lapidus. Michigan State University, East Lansing, MI, USA.

Though the wormlike chain (WLC) model has successfully described the statistical properties of fully denatured polypeptides, the lack of sequence details and attractive forces made it less successful in describing unfolded states in folding conditions. To cover the limitation while keeping the model's efficient feature, we have developed a sequence specific wormlike chain model. Computationally, secondary structure constraints from the secondary chemical shift measurements of drkN are integrated into the construction of each wormlike chain. Then the probability distribution is reweighted to bias compact conformations in which residues of similar hydrophobicity are located near each other. This model has been tested on two mutants (C2 and C60) of the Drosophila drk N-terminal (drkN) SH3 domain. drkN exists in approximately 1:1 equilibrium between folded and unfolded state in water, which gives us the opportunity to monitor the contact quenching of tryptophan 36 by either cysteine under all denaturing conditions. The experimental results exhibit a relatively slow kinetics, which implies slow intramolecular diffusion. The reweighted pairwise distance distributions are also compared to the Paramagnetic Relaxation Enhancement (PRE) data

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for drkN.

Comparison of λ Cro Solution Ensemble To Its Open and Closed Crystal Forms

Logan S. Ahlstrom, Osamu Miyashita.

University of Arizona, Tucson, AZ, USA.

λ Cro is a small dimeric bacteriophage transcription factor consisting of multiple crystal structures. Its X-ray images range from a more closed DNA-free conformation relative to an open form when bound to DNA. Considered a flexible "dumbbell" with a pliable β-sheet region connecting two more rigid αhelical domains, the subunits of λ Cro may undergo a relative rotation. This suggests a possible induced fit DNA-binding model (1). In solution the protein likely accesses a range of conformations encompassing its DNA-bound open and DNA-free closed forms. However, the snap-shots from X-ray crystallography capture just one of these structures, making it difficult to recompose the λ Cro solution ensemble. Moreover, the crystal may have randomly selected a conformer un-representative of its dominant physiological form. Here we show the transition between the open and closed forms of λ Cro to occur with no large energy barrier in solution. We have performed 50 ns all-atom Molecular Dynamics (MD) simulations starting from both structures in explicit solvent using the Amber10 force field. The trajectory started from the closed form undergoes transitions to a structure intermediate of the open and closed conformations. The simulation begun from the open form closes spontaneously. We have extended our work in a Generalized Born continuum model as a basis for enhanced sampling techniques to obtain the free energy landscape of λ Cro conformational change. This will provide insight into the protein's DNA recognition mechanism. We anticipate our solution simulations to be the starting point to study the effects of crystal environments on λ Cro conformation (2). [1] B. M. Hall, et al. (2008) J. Mol. Biol. 375, 802-811. [2] I. I. Vorontsov and O. Miyashita (2009) Biophys. J. doi:10.1016/j.bpj.2009.08.011

Protein Folding & Stability I

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Rheology of Small Volume Antibody Formulations and Kinetics of Surface Induced Protein Fibrillation

Matthew C. Dixon.

Q-Sense, Glen Burnie, MD, USA.

This study shows how the QCM-D technology can shed light onto complex protein-protein interactions by rheological characterization of high concentration antibody solutions and measuring amyloid growth kinetics. Quartz Crystal Microbalance with Dissipation (QCM-D) is a surface sensitive technique which simultaneously measures the mass and viscoelastic properties of adsorbed layers and bulk fluids. Bulk fluid characterization was demonstrated by varying the pH of a humanized monoclonal antibody in 10mM sodium acetate or MES buffer and flowing ~ 350 ul of this solution at a rate of 50 ul/min across a QCM-D sensor. The storage (G') and loss moduli (G") of the IgG2 antibodies were measured between a pH range of 4 - 6.2 at a variety of different frequencies. The ratio of G"/G' was greater than 1 for all solutions investigated with a maximum found at pH 5.5 indicating the greatest liquid-like behavior relevant both to administration and also physical characterization of the formulation itself. Surface fibrillation was studied by anchoring an insulin seed fibril onto an inert QCM-D surface and measuring the uptake of additional insulin as a function of time that was later correlated to AFM images of the surface. Growth kinetics of insulin were measured by QCM-D onto seed surfaces and found to be 83 +/-0.2 pg/s or $8.1 \times 10^9 + -1.4 \times 10^6$ individual insulin molecules per second. In the absence of seed fibrils insulin uptake was negligible. The rates of fibril growth accurately follow Arrhenius type behavior with an enthalpic activation barrier of ΔH^{\ddagger} = 24.4 +/- 1 kcal/mol and a Gibbs free energy of 6.1 +/- 2 kcal/mol.

151-Po:

Direct Observation of the Tug-Of-War During the Folding of a Mutually Exclusive Protein

Qing Peng, Hongbin Li.

University of British Columbia, Vancouver, BC, Canada.

Although most protein folding studies are carried out on single-domain proteins, over two thirds of proteins in proteomes are multi-domain proteins. A significant fraction of these multi-domain proteins are domain-insertion proteins, in which one guest domain is inserted into a surface loop of a host protein. Intricate thermodynamic and kinetic coupling between the two domains can have profound impacts on their folding dynamics, which has not been probed yet. Here we engineered a mutually exclusive protein as a model system to directly illustrate one such complex dynamic process: the 'tug-of-war' process during protein folding. By inserting a guest protein I27w34f into a host protein GB1-L5 (GL5), we engineered a novel mutually-exclusive protein GL5/ 127w34f, in which only one domain can remain folded at any given time due to topological constraints imposed by the folded structures. Using stoppedflow techniques, we obtained the first kinetic evidence that the guest and host domains engage in a folding tug-of-war as they attempt to fold, in which the host domain folds rapidly into its three-dimensional structure and is then automatically unfolded driven by the folding of the guest domain. Our results provided direct evidence that protein folding can generate sufficient mechanical strain to unravel a host protein. Using single molecule atomic force microscopy, we provide direct evidence for the existence of a conformational equilibrium between the two mutually exclusive conformations. Our results highlight important roles played by the intricate coupling between folding kinetics, thermodynamic stability and mechanical strain in the folding of complex multi-domain proteins, which cannot be addressed in traditional singledomain protein folding studies.

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Protein Oligomerization: Thermodynamic and Structural Analysis of the Dimerization of Beta-lactoglobulin

Martha I. Burgos, Sergio A. Dassie, Gerardo D. Fidelio.

School of Chemistry, National University of Córdoba, Córdoba, Argentina. Protein oligomers are widely found in living organisms and they are also crucial in some neurodegenerative diseases. The aim of this work is to analyze the thermodynamics and stability of protein oligomers employing β -lactoglobulin as the experimental model.

 β -lactoglobulin is a small, globular and compact protein which, in solution, is in equilibrium with its dimeric form. The association equilibrium can be affected through changes in total protein concentration, temperature and pH. The dimer is the prevalent species at neutral pH. We studied the dissociation process of β -lactoglobulin induced by temperature employing different techniques at pH = 6.7. We performed calorimetric experiments at different values of total protein concentration and we observed that the endotherms of